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Mechanism of Squalene Cyclization: The Chiral Origin of the C-22 Hydrogen Atoms of Fusidic Acid

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Summary The C-2 protons of mevalonic acid are incorporated with retention of configuration into C-22 of fusidic acid; this finding excludes the intermediacy of products with a $\Delta^{20(22)}$ double bond in the formation of a $\Delta^{17(20)}$ double bond in fusidic acid.

The difficulty^{1,2} of rationalizing the collapse of cation (1)³ to protosterol antibiotics like fusidic acid (2a) and others⁴ all having the Z geometry at $\Delta^{17(20)}$ has been pointed out. Stabilization of (1) through direct elimination of the 17β -proton (route 'a') would lead according to Cornforth's hypothesis^{3b} to the wrong geometry about the 17(20) double bond.^{1,2} Additionally in such a process the cyclase system of F. coccineum would be expected to obviate the backbone rearrangement³ through a kinetically controlled stabilization of (1) (route 'a'). In contrast during the enzymatic transformation of (1) into lanosterol, it has been suggested that the cyl se system exerts only a marginal influence.⁵

The interpretational difficulty could be alleviated were the formation of (2a) to proceed through a $\Delta^{20(22)}$ intermediate (3a) (1; route 'b') as noted by Corey et al.⁶ for the analogue (3b). Enzymatic isomerization of (3a) could afford (2a) (origin of C-22 hydrogens not assigned). Rotation around C-17(20) of (3a) would permit positioning of the side chain and the 17-hydrogen suitable for the formation of a $\Delta^{17(20)}$ double bond in (2a).

The presence of a $\Delta^{20(22)}$ intermediate was supported by an observation that incubation^{1,7} of (3RS;2S)-[2-1⁴C,2-³H]-mevalonic acid (MVA) (³H: ¹⁴C ratio 5·26: 1 atomic ratio 1: 1) with F. coccineum gave S-fusidic acid (2a) with a ³H: ¹⁴C ratio of 4·65: 1, corresponding to the incorporation of 5·3 atoms of tritium and 6 atoms of ¹⁴C. In the case of R-fusidic acid (2a) biosynthesized from (3RS,2R)-[2-1⁴C,2-³H]-MVA (³H: ¹⁴C ratio 5·15: 1) the ³H: ¹⁴C ratio of 5·04: 1 corresponded to an atomic ratio of 6: 6. The results with (2S)-[2-1⁴C,2-³H]-MVA could be interpreted as arising from the

II; (2S)-[2-3H,2-14C]MVA

I; (2R)-[2-8H,2-14C]MVA

Experiment with:

	•		Derivative	¹⁴ C-Spec,	3H:14C ratio		¹⁴ C-Spec. act. ⁸	8H:14C ratio	
No	No. Product		counted	act.	Isotopic	Atomic		Isotopic	Atomic
l.	Mevalonic acid	• •	N-Diphenyl- methylamide	8.79	5.15	1.02:1	4.75	$5 \cdot 26$	1.13:1
2.	Methyl fusidate		(2b)	86.7	5.04	6.00:6	$82 \cdot 4$	4.65	6.00:6
3.	Methyl dihydrofusidate		(2c)	85.6	5.05	6.00:6	82.5	$4 \cdot 62$	5.96:6
4.	6-Methylheptane-1,2-diol		(4b)	30.2	4.92	1.95:2	29.0	4.58	1.97:2
5 .	5-Methylhexan-1-al		(4d)	30.4	5.04	2.00:2	$29 \cdot 0$	4.66	2.00:2
6.	4-Methylpentan-1-ol		(5c)	$2 \cdot 7$	5.02	1.99:2	4.0	4.60	1.98:2
7.	4-Methylpentan-1-al		(6b)	1.5	3.02	1.20:2	4.9	4.54	1.95:2
8.	Methyl 4-methylpentanoate	• •	(6d)	_	_	_		2.38	1.02:2

^a D.p.m. per mmol \times 10⁴; entries 1, 6 and 7 were counted at higher dilutions.

loss of a 22-pro-S hydrogen via pathway ['b' shown in (1)] in the biosynthesis of (2a). The specimens of R and S-fusidic acid were thus degraded as shown in the Scheme and the chirality and ³H content at C-22 determined.

● - C-2 carbon of MVA

 H_R and H_S — protons from the 2-pro-R and 2-pro-S of MVA \bigoplus — protons from 4-pro-R of MVA.

The products† obtained from R-fusidic (Table; Experiment I; entries 2—6) exhibited 3H : ${}^{14}C$ ratios which corresponded to the predicted 1,8 atomic ratios. One atom of tritium‡ was lost on the NAD+-YADH oxidation of (5b) to (6a) (Experiment I; entries 6, 7), thus establishing the 1R configuration of the alcohol. 11

The products from the S-fusidic acid had constant ³H: ¹⁴C ratios (Experiment II; entries 2-7). Clearly NAD+-YADH

oxidation of the alcohol (5b) to aldehyde (6a) proceeded without loss of tritium. Hence if (5b) had a tritium atom at C-1 the alcohol must have the 1S configuration. Consequently the diene^{1,12} (7) (from S-fusidic acid) was oxidized¹³ to (6c) which was purified as the methyl ester (6d) by g.l.c. The ³H: ¹⁴C ratio of (6d) indicates that of the two ³H atoms present in (5b) only one was retained. Therefore

a; i $\rm H_2;^{7.9}$ ii $\rm CH_2N_2.$ b; i $\rm O_3;$ ii $\rm LiAlH_4.$ c; $\rm H_5IO_6.$ d; i $\rm CF_5-COOOH;^{10b}$ ii $\rm LiAlH_4;$ iii prep. g.l.c. e; NAD+-Yeast alcohol dehydrogenase (YADH). f; i $\rm LiCl-HCO\cdot NMe_2;$ 12 ii $\rm CH_2N_2.$ g; i $\rm RuO_4;$ 13 ii $\rm CH_2N_2;$ iii prep. g.l.c.

one atom of tritium must be present at C-1 of (5b) and (6a). It follows that a tritium atom originating from (2S)-[2-14C,-2-3H]-MVA is present a the 22-pro-S position of the derived S-fusidic acid.

The results demonstrate that the C-2 protons of MVA are incorporated into C-22 of fusidic acid with retention of their stereochemical integrity. The intermediacy of a $\Delta^{20(22)}$

† All the compounds were homogenous by g.l.c. or t.l.c. Derivatives were identified by n.m.r., mass, and i.r. spectroscopy, etc., and by comparison with authentic samples.

‡ The removal of only 0.8 atom of tritium may have been caused by partial air oxidation of the alcohol (see ref. 11).

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precursor is thus excluded. Because the C-13 hydrogen of fusidic acid is derived from the 4-pro R proton of MVA,1 Δ^{12} or $\Delta^{13(17)}$ intermediates also cannot be involved in the biosynthesis. Hence it seems that the stabilization of (1) in the formation of (2a) cannot be rationalized solely on an organic chemical basis⁵ without some enzyme participation.¹

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- I. J. Mulheirn and E. Caspi, J. Biol. Chem., 1971, 246, 2494; E. Caspi and L. J. Mulheirn, J. Amer. Chem. Soc., 1970, 92, 404.
 I. J. Mulheirn and P. J. Ramm, Chem. Soc. Rev., 1972, 1, 259.
 (a) A. Eschenner, L. Ruzicka, O. Jeger, and D. Arigoni, Helv. Chim. Acta, 1955, 38, 1890; (b) J. W. Cornforth, Angew. Chem. Internat. Edn. 1968, 7, 903.
- (a) T. G. Halsall, E. R. H. Jones, G. Lowe, and C. E. Newall, Chem. Comm., 1966, 685; P. Oxley, ibid., p. 729; (b) S. Okuda, S. Iwasaki, M. I. Sair, Y. Machida, A. Inoue, K. Tsuda, and Y. Nakayama, Tetrahedron Letters, 1967, 2295; W. von Daehne, H. Lorch, and W. O. Godtfredsen, ibid., 1968, 4843.

- ⁵ E. E. van Tamelen and J. H. Freed, J. Amer. Chem. Soc., 1970, 92, 7206; E. E. van Tamelen, R. P. Hanzlik, R. B. Clayton, and A. L. Burlingame, *ibid.*, p. 2137.

 ⁶ E. J. Corey, P. R. O. de Montellano, and H. Yamamoto, J. Amer. Chem. Soc., 1968, 90, 6255; E. J. Corey, K. Lin, and H. Yamamto, *ibid.*, 1969, 91, 2132.
 - ⁷ W. O. Godtfredsen, W. von Daehne, S. Vangedal, A. Marquet, D. Arigoni, and A. Melera, Tetrahedron, 1965, 21, 3505.

 - ⁸ G. Popjak and J. W. Cornforth, *Biochem. J.*, 1966, **101**, 553. ⁹ W. O. Godtfredsen and S. Vangedal, *Tetrahedron*, 1962, **18**, 1029.
- ¹⁰ (a) J. B. Greig, K. R. Varma, and E. Caspi, J. Amer. Chem. Soc., 1971, 93, 760; (b) M. F. Hawthorne, W. D. Emmons, and K. S. McCallum, ibid., 1958, 80, 6393.
- ¹¹ J. B. Greig, K. R. Varma, and E. Caspi, J. Amer. Chem. Soc., 1971, 93, 760; E. Caspi, K. R. Varma, and J. B. Greig, Chem. Comm., 1969, 45; K. R. Varma and E. Caspi, J. Org. Chem., 1969, 34, 2489.
 W. O. Godtfredsen, W. von Daehne, and S. Vangedal, Chem. Comm., 1966, 638.
- ¹⁸ D. M. Piatak, H. B. Bhat, and E. Caspi, J. Org. Chem., 1969, 34, 112; D. M. Piatak, G. Herbst, J. Wicha, and E. Caspi, ibid., p. 116.